The Total Synthesis of Proteasome Inhibitors TMC-95A and TMC-95B: Discovery of a New Method To Generate *cis*-Propenyl Amides**

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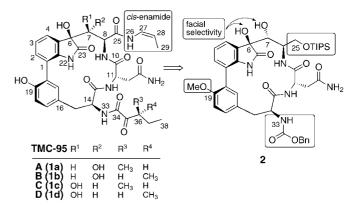
The ubiquitin proteasome pathway is critical for accomplishing proteolysis in both the cytosol and nucleus of all eukaryotic cells. Understanding of the physiological roles of a particular proteasome has been aided by studying the effects of cell-permeable inhibitors. Moreover, specific proteasome inhibitors are emerging as possible drug candidates. Let Accordingly, the discovery of the cyclic peptides TMC-95A-D (1a-d; see Scheme 1), first isolated as fermentation products of *Apoispora montagnei* SACC TC 1093 derived from soil samples, which differ in stereochemistry at the remote, configuratively labile C36 center, inhibit proteasomal functions of the 20S proteasome with IC₅₀ values down to low nanomolar levels. [2a]

Accordingly, a total synthesis program, which targets the more active TMC-95A and B isomers, was launched in our laboratory. We hoped to gain access to both compounds since after a successful total synthesis the groundwork would be more promising for establishing systematic SAR profiles of these TMC-95 inhibitors. Ideally, simpler structures could be designed, synthesized, and screened for similar activity in anticipation of their use as new drug leads.

Recently, we described an approach to accomplish stage I in our program, that is, the total synthesis of the TMC-95 inhibitors. [4a] The most advanced structure reported in our previous disclosure was compound 2 (Scheme 1). The attainment of this subgoal, promising as it was for our proposed program, still left unaddressed several key issues for total syntheses of 1a and 1b. Thus, provision would be necessary to allow exposure of a free phenolic hydroxy group at C19 and a homopyruvoyl group at N33 (see structure 2). Moreover, the oxidation level at C25 must be upgraded from an alcohol, as found in 2, to the carboxy state required to reach 1 (see below). This was no trivial matter, given the free hydroxy groups at C6 and C7 in our model target, 2. We also note that in the synthesis of 2 the facial sense of *cis* dihydroxylation at

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Scheme 1. Structures of TMC-95A-D (1a-d) and 2. TIPS = triisopropylsilyl, Bn = benzyl.

C6 and C7 was virtually stereorandom. [4a] Perhaps most problematic was the *cis*-propenyl amide linkage that encompasses atoms N26–C29 of the multifaceted molecular ensemble of **1**. It is with this latter issue that we commence our report.

Examination of the literature reveals several methods that might, in principle, be relevant to the enamide problem.^[5] However, the applicability of these methods to the synthesis of the active TMC compounds is not certain. Compounds 1 have a rich diversity of functionality, particularly in the extended "pyruvoyl"-like (C34-C38) and dihydroxyindolinone (positions 22, 23, and 6-8) regions. Anticipation of potential vulnerability in these sectors, not to speak of the cisenamide itself (N26-C29), prompted us to explore a new modality for reaching such a substructure appropriate for molecules with multiple sites of potential instability. We wondered whether a substance of the type 3 might, upon heating or catalysis, undergo concurrent ene- and silatropiclike bond reorganizations that would lead to 4 (Scheme 2). If this hypothesis were to be fruitful, the cis character of the enamide would be virtually assured at the kinetic level. Moreover, it seemed at least possible that the intermediate silyl imidate linkage in 4 could be cleaved, with retrieval of general substructure 5.

Scheme 2. Formation of *cis*-propenyl amides **5** by rearrangement – hydrolysis of α -silylallyl amides **3** (R = aryl, alkenyl, and alkyl groups).

In the event, a range of probe substrates **3** was synthesized by acylation of the known of amine **6** (Table 1).^[6] As seen in entries 1–3, heating of these compounds at about 110 °C for the time periods indicated gave rise to silyl imidates **4a–c** (observed by ¹H NMR analysis).^[7] Aqueous hydrolysis of these compounds afforded enamides **5a–c**. The reaction was also applicable to substrate **3d** (entry 4), though a longer heating time was required for its conversion to **4d**. Most significantly, the rearrangement – hydrolysis sequence proved extendable to the aminoacyl substrate **3e** (entry 5) to afford

Tabelle 1. Rearrangement – hydrolysis of α -silylallyl amides 3.[a]

TES RCOCI, NEts; or RCO₂H, EDC/HOAT R NH NH ROPE Single separate
$$\mathbf{6}$$
 82-95% $\mathbf{3}$ $\mathbf{5}$ single separate $\mathbf{6}$

		130	nii Gi	
Entry	R =	Conditions	Yield [%]	
1	a C	a) toluene, 110 °C, 10 h; b) H_2O	81	
2	p .0	a) toluene, 110 °C, 20 h; b) H_2O	73	
3	c ()	a) toluene, 110°C, 27 h; b) H ₂ O	67	
4	d \\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	a) toluene, 110° C, 3 d; b) H_2 O	72	
5	e TIPSO Th. Boc	a) o-xylene, 135° C, $4\mathrm{d}$; b) $\mathrm{H}_2\mathrm{O}$	52	

[a] TES = triethylsilyl, Boc = *tert*-butoxycarbonyl, EDC = 1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride, HOAT = 1-hydroxy-7-azabenzotriazole.

4e and thence **5e**. It was of great interest to determine whether this new method would find application at a very late stage of our projected total synthesis.

The starting material for our refashioned synthesis was the previously described iodooxindole 7 (Scheme 3). [4a] Crossed-aldol condensation of 7 with the Garner aldehyde 8, [8] followed by β -elimination of the derived mesylate, afforded

Scheme 3. Synthesis of biaryl compound **14.** a) LDA (2.0 equiv), THF, $-78\,^{\circ}$ C, 1.5 h; NEt₃, MsCl, CH₂Cl₂, $-70 \rightarrow -50\,^{\circ}$ C, 1.5 h; 81% (E/Z=1.3:1); b) I₂ (cat.), benzene, 80°C, 26 h; DMP/PPTS, toluene, 65°C, 5 h; 85% (60% conversion); c) 1) MeOH/SOCl₂; 2) CbzCl/K₂CO₃; 3) BnBr, Cs₂CO₃, acetone, reflux; 88% (three steps); d) Ag₂SO₄/I₂, MeOH, RT, 1 h; 99%; e) pinacolatodiborane, [PdCl₂(dppf)] · CH₂Cl₂, KOAc, DMSO, 80°C, 10 h, 91%; f) (E)-**9**, [PdCl₂(dppf)] · CH₂Cl₂, K₂CO₃, DME, 80°C, 2 h; 75%. Cbz = benzyloxycarbonyl, LDA = lithium diisopropylamide, DMP = 2,2-dimethoxypropane, PPTS = pyridinium p-toluenesulfonate, dppf = bis(diphenylphosphanyl)ferrocene.

a 1:1.3 mixture of $\alpha.\beta$ -unsaturated lactams (Z)-9 and (E)-9. [^{4a,e]} Fortunately, the former isomer could be converted to the latter one through iodine-mediated isomerization. [^{4a,e]} In a parallel sequence, L-tyrosine (**10**) was converted to **11** in three steps (Scheme 3). [9] A high-yielding *ortho* iodination [^{4a]} of **11** led to **12** and thence, after palladium-mediated borylation, [10] to **13**. Suzuki-type coupling [11] of **13** with (E)-9 afforded compound **14** (75% yield). We hoped that the biaryl domain thus presented would be serviceable in the context of our projected total synthesis (see below).

Hydrolysis of the methyl ester function of **14** led to the corresponding carboxylic acid, which, after acylation of the basic nitrogen atom of the asparagine derivative H-Asn-OtBu (**15**), afforded **16**. The hydroxy groups were introduced at C6 and C7, as shown in Scheme 4. Indeed, the presence of the Garner N,O-acetonide served to direct, preferentially, the oxidizing agent to the Re face (C6) of **16** to afford **17** in a 5:1 ratio relative to its 6R,7S stereoisomer (not shown).

Scheme 4. Synthesis of diol **17**. a) 1) LiOH, THF/H₂O, 0° C, 1.5 h; 2) H-Asn-O*t*Bu (**15**), EDC/HOAT, THF, RT, 2 h; 85 % (two steps); b) OsO₄/NMO, (DHQD)₂-PHAL, *t*BuOH/H₂O, RT, 12 h; 88 % (dr = 5:1). Asn = asparagine, NMO = 4-methylmorpholine *N*-oxide, (DHQD)₂-PHAL = 1,4-bis(9-*O*-dihydroquinidine)phthalazine.

The timing and manner in which the various heteroatomcentered functional groups were exposed and protected proved to be critical to the success of the project. We proceeded as follows (see Scheme 5): Deprotection of the N,O-acetonide linkage of 17 afforded N-Boc triol 18. The primary alcohol at position 25 was protected as its TIPS derivative (19). Cleavage of the tert-butyl group generated a carboxylic acid at C10 which set the stage for macrolactamization (see compound 20).[12] In the next step, the benzyl groups protecting the C19 phenol and N33 were concurrently cleaved by hydrogenolysis. Acylation of the latter basic nitrogen atom with racemic 21[13] afforded 22a and 22b as a 1:1 mixture. Both compounds were advanced concurrently. The TIPS protecting group was cleaved from the primary alcohol. The four hydroxy groups (positions 6, 7, 19, and 25) were protected as the TES derivatives (see 23a and 23b). In a key step of the synthesis, reaction of these compounds with Jones reagent^[14] led to specific oxidation at the primary center (position 25) to afford acids 24a and 24b. In addition, partial deprotection occurred at the C19 ether linkage which resulted in the formation of 25a and 25b. This four-component mixture led, after condensation with amine 6 as indicated, to the amide silvl ethers **26a** and **26b** as well as amide phenols **27a** and **27b** (**26a,b**:**27a,b** \approx 1:1.5).

Scheme 5. Synthesis of α -silylallyl amides **26a,b** and **27a,b**. a) PPTS/MeOH, reflux, 2 h; b) TIPSCl, imidazole/DMAP, CH₂Cl₂, RT, 5 h; 88% (two steps); c) 1) TFA/CH₂Cl₂ (4:1), RT, 2 h; 2) EDC/HOAT/DIEA, CH₂Cl₂/DMF(2 mM), RT, 24 h; 52% (two steps); d) 1) Pd/C, H₂, EtOH, RT, 19 h; 2) (\pm)-3-methyl-2-oxopentanoic acid (**21**), EDC/HOAT, CH₂Cl₂/DMF, RT, 2 h; 85% (2 steps); e) 1) HF/Py; 2) TESOTf, 2,6-lutidine, CH₂Cl₂, 0°C \rightarrow RT, 15 h; 3) NaHCO₃; 4) citric acid, EtOAc/H₂O; 73% (from **22**); f) Jones reagent, acetone, 0°C, 2 h; g) **6**, EDC/HOAT, CH₂Cl₂/DMF, RT, 13 h; 45% (two steps). DMAP = 4-dimethylaminopyridine, TFA = trifluoroacetic acid, DIEA = *N*,*N*-diisopropylethyl amine.

Construction of the (Z)-1-propenylamide was now achieved by thermally driven rearrangement of the α -silylallyl amides corresponding to 3 (Scheme 6). The rearrangement of

Scheme 6. Synthesis of TMC-95A (1a) and TMC-95B (1b). a) 1) o-xylene, 140 °C, 3 d; 2) H₂O; b) HF/Py, THF/Py; then Me₃SiOMe; 49% (two steps).

the complex mixture^[15] in anhydrous o-xylene at $140\,^{\circ}$ C provided (Z)-1-propenylamides $28\,a$,b and $29\,a$,b. The crude mixture of these compounds was globally deprotected with pyridine-buffered HF/pyridine to afford a mixture of our total synthesis goals—TMC-95A and TMC-95B (1a and 1b; 1:1). This mixture was separated by RP-HPLC^[16] to provide the individual compounds 1a and 1b. These were characterized

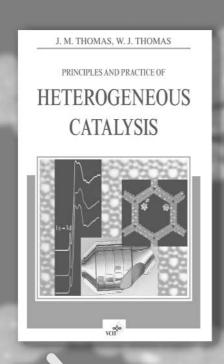
by their high-field NMR spectra in comparison with those of authentic samples.^[16]

In summary, the total syntheses of TMC-95A and TMC-95B have been achieved. The program featured a sequential assembly of oxindole 7, Garner's aldehyde (8), aryl boronate 13, asparagine derivative 15, 3-methyl-2-oxopentanoic acid (21), and α -silylallyl amine 6. Highlights of the synthesis include a Suzuki biaryl construction ((E)-9 + 13 \rightarrow 14), a diastereofacial dihydroxylation reaction that took advantage of the Garner method (16 \rightarrow 17), and a macrolactamization (formation of 20). We also note that new chemistry to accomplish stereospecific *cis*-propenyl amide formation (26/27 \rightarrow 28/29) was inspired by goal system 1. The application to the delicate case at hand serves to build confidence in its generality.

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- [6] Amine 6 (S.-F. Chen, E. Ho, P. S. Mariano, Tetrahedron 1988, 44, 7013-7026) was synthesized by using an improved procedure from allyl alcohol through a one-pot TES ether formation, retro-Brook rearrangement, and mesylation, followed by a displacement of mesylate with ammonia. Acylation in entries 1-4 (Table 1) was accomplished by coupling of 6 with the acid chlorides, while in entry 5 an EDC-mediated coupling with protected amino acid with amine 6 was involved. Details will be forthcoming in a full disclosure.
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- [12] The atropisomer shown for **20** follows from the C6 stereochemistry (see ref. [2b]).

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- separation was achieved at the stage of the two-component mixture 1a/1b
- [16] HPLC conditions are as follows. Column: YMC-pack ODS-AM, 150×10 mm; eluant: 25 % MeCN in water; flow rate: 2.5 mL min⁻¹, $t_{\rm R}({\bf 1a})=37.3$ min, $t_{\rm R}({\bf 1b})=34.3$ min. The synthetic materials were identical to natural TMC-95A and TMC-95B (TLC, NMR, EIMS, and HPLC). We thank Dr. Jun Kohno, Tanabe Seiyaku Co., Japan, for generously providing us with the HPLC conditions and the samples of TMC-95A and B for comparison.



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